

Specimen

C.difficile Toxin Assay

- › Unformed stool collected in clean plastic screw-cap container.

Enteric Pathogen Assay

- › Stool – 1 gram collected in Para-Pak C & S (Orange Cap Vial) for Enteric Pathogen Assay
- › If multiple stool cultures are requested, they must be collected at least 24 hours apart.

Giardia/ Cryptosporidium Screen

- › Stool in SAF fixative (pink O&P vial)
- › Recommended collection: Three separate stool specimens within a 10-day period, each at least 24 hours apart.

Ova and Parasite

- › Stool in SAF fixative (green O&P vial)
- › Stool in 10% formalin preservative
- › Recommended collection: Three separate stool specimens within a 10-day period, each at least 24 hours apart.

Pathogens Tested For

- › The Enteric Pathogen Assay includes: Campylobacter, E. coli (STEC) Shiga toxin 1 and Shiga toxin 2, Salmonella species, Shigella species, Vibrio, Yersinia enterocolitica, Norovirus, and Rotavirus
- › For Aeromonas or Pleisiomonas orders, add on “Culture- Aeromonas/Pleisiomonas.”
- › For Cyclospora, collect stool specimen as for ova and parasites and add comment “Suspecting Cyclospora.”
- › For Isospora, collect stool specimen as for ova and parasites and add comment “Suspecting Isospora.”

Unacceptable Specimens

- › Delay greater than 2 days (48 hours) in transport of preserved specimen using *Para-Pak C & S* stool transport vial (orange cap vial).
- › Only one sample will be tested if two or more are collected within 24 hours of each other.
- › Specimens contaminated with urine
- › Specimens not properly labeled.
- › For C.diff testing: Formed stools, frozen stools, specimens collected within 7 days of a previously tested sample or with a previous positive resulted in the past 30 days

Material Needed

Obtain collection kits from [North Memorial Health Laboratory](#). Once added, specimen is stable up to 48 hours.

Stool Specimen Collection:

1. If patient is taking antacids, antidiarrheal medication or oily laxatives please attach a note to the specimen stating so. Specimens should not be collected for 7 to 10 days after barium or bismuth has been given.
2. When more than one specimen is requested, space collection at 2 or 3 day intervals. Specimens must be at least 24 hours apart.
3. Pass the stool onto a clean dry surface, such as a bedpan, a clean margarine tub, a clean wide mouthed jar or a clean milk carton with the top cut off. A large plastic bag placed into a wastebasket to catch the specimen can also be used. Do not take the specimen from the toilet or contaminant with urine.
4. Open the vial containing the liquid. Using the collection spoon built into the lid, place small scoopfuls of stool from areas which appear bloody, slimy or watery into the vial until the contents rise to the **red line**. If the stool is formed (hard), please try to sample small amounts from each end and the middle.

5. Mix the stool on the sides of the vial with the spoon, then twist the cap tightly closed and shake the vial vigorously until the contents are well mixed. Wipe off the outside of the vial if any spillage occurs. CHECK THE CAP TO BE CERTAIN IT IS TIGHTLY CLOSED.
6. Label both the vial and the bag with the patient name, and date and time collected. Place the vial back into the bag. Do NOT cover this information with another label.
7. Keep the specimen(s) at room temperature until they can be returned to the laboratory. Specimens should be returned to the laboratory with 72 hours of their collection. If more than one O&P specimen is requested, it may be advantageous to return the first one collected as soon as possible so testing can begin, rather than waiting until all have been collected.